DOI: 10.1002/cmdc.200800257

Apoptosis-Inducing High 'NO Concentrations Are Not Sustained Either in Nascent or in Developed Cancers

Adam Heller*[a]

Nitric oxide (NO) induces apoptosis at high concentrations by Snitrosating proteins such as glyceraldehyde-3-phosphate dehydrogenase. This literature analysis revealed that failure to sustain high 'NO concentrations is common to all cancers. In cervical, gastric, colorectal, breast, and lung cancer, the cause of this failure is the inadequate expression of inducible nitric oxide synthase (iNOS), resulting from the inhibition of iNOS expression by TGF- β 1 at the mRNA level. In bladder, renal, and prostate cancer, the reason for the insufficient 'NO levels is the depletion of arginine, resulting from arginase overexpression. Arginase competes with iNOS for arginine, catalyzing its hydrolysis to ornithine and urea. In gliomas and ovarian sarcomas, low 'NO levels are caused by inhibition of iNOS by N-chlorotaurine, produced by infiltrating neutrophils. Stimulated neutrophils express myeloperoxidase, catalyzing H_2O_2 oxidation of Cl⁻ to HOCl, which N-chlorinates taur-

A common cause of cancers: failure to induce apoptosis by maintaining high **NO concentrations**

At high concentrations ($>10^{-6}$ m), nitric oxide ('NO) induces apoptosis by S-nitrosating glyceraldehyde-3-phosphate dehydrogenase (GAPDH) $^{[1-3]}$ and other proteins. This analysis of 195 publications indicates that a failure to attain high 'NO concentrations is characteristic of all cancers. In cases of cervical, $^{[4]}$ gastric, $^{[5]}$ colorectal,^[6] breast,^[7,8] and $\text{lung}^{[9]}$ cancers, the reason for the inadequate level of 'NO is the insufficient expression of inducible nitric oxide synthase (iNOS), resulting from TGF- β 1-mediated inhibition of iNOS expression at the mRNA level.[10] Additionally, in some cases of bladder,^[11] renal,^[12,13] and prostate^[11,14,15] cancer, the cause of failure is depletion of arginine, resulting from arginase overexpression. Arginase competes with iNOS for arginine, catalyzing its hydrolysis to orni-

[a] Prof. A. Heller Department of Chemical Engineering University of Texas Austin, TX 78712 (1154) Fax: (+1) 512-471-8799 E-mail: heller@che.utexas.edu

thine and urea. In certain gliomas $[16, 17]$ and ovarian sarcomas, $^{[18]}$ inhibition of iNOS by N-chlorotaurine (taurine chloramine) leads to decreased 'NO levels.^[19] Stimulated neutrophils express myeloperoxidase,^[20, 21] which catalyzes H_2O_2 mediated oxidation of CI^- to HOCl, which in turn N-chlorinates taurine^[22,23] at the cellular taurine concentration of 19 $\text{mm}^{[24]}$ in neutrophils. In cases of squamous cell carcinoma of the skin,^[25] ovarian cancer,^[26] lymphoma,^[26] Hodgkin's disease,^[26] and breast cancer,^[27] Nbromotaurine (taurine bromamine) inhibits iNOS through a similar mechanism. N-bromotaurine, produced by eosinophil-peroxidase-expressing infiltrating eosinophils,^[28] is formed by the oxidation of Br^- to HOBr by H_2O_2 , which in turn Nbrominates taurine to N-bromotaurine^[29, 30] at a cellular concentration of 15 mм in eosinophils.^[24]

Circulating red blood cells deplete tumoral 'NO levels and arrest 'NOinduced apoptosis

Nitric oxide S-nitrosates red blood cell (RBC) glutathione^[31] and hemoglobin (Hb) , $[32, 33]$ and is rapidly oxidized to ni-

ine at its concentration of 19 mm in neutrophils. In squamous cell carcinomas of the skin, ovarian cancers, lymphomas, Hodgkin's disease, and breast cancers, low 'NO concentrations arise from the inhibition of iNOS by N-bromotaurine, produced by eosinophil-peroxidase-expressing infiltrating eosinophils. Eosinophil peroxidase catalyzes the H₂O₂ oxidation of Br⁻ to HOBr, which Nbrominates taurine to N-bromotaurine at its concentration of 15 mm in eosinophils. In microvascularized tumors, the 'NO concentration is further depleted; 'NO is rapidly consumed by red blood cells (RBCs) through S-nitrosation of RBC glutathione and hemoglobin, and by oxidation to nitrate by RBC oxyhemoglobin. Angiogenesis-inhibiting antibodies are currently used to treat cancers; their mode of action is not, as previously thought, reduction of the tumor $O₂$ or nutrient supply. They actually decrease the loss of 'NO to RBCs.

> trate by oxyhemoglobin. $[34]$ The three reactions decrease the concentration of 'NO, decrease apoptosis, and increase cancer virulence. Angiogenesis-inhibiting antibodies currently used in the treatment of cancers are thought to limit the supply of $O₂$ or nutrient levels in the tumor: $[35-40]$ however, they actually decrease the 'NO loss to the RBCs in the microvasculature.

> High concentrations of 'NO are reached when tumor stromal cells $[41]$ and infiltrating macrophages are stimulated to express iNOS, which catalyzes the $O₂$ mediated oxidation of arginine to 'NO and citrulline. Stimulated iNOS-expressing macrophage cultures sustain the highest reported concentrations of 'NO, as high as 2×10^{-4} m. $^{[42]}$ In contrast, the highest reported neuronal nitric oxide synthase (nNOS)-sustained 'NO concentrations are 10^{-6} m, ^[43] and endothelial nitric oxide synthase (eNOS)-sustained "NO concentrations are only 2×10^{-8} M.^[44] iNOS is expressed in most, but not in all, cancers.

Daughter products of 'NO are mutagenic,^[45] and at low concentrations, 'NO induces VEGF-mediated cancer-aggravating angiogenesis.^[41,46-50] For these reasons, many researchers view 'NO as a cause of cancer, rather than as a longrange cancer-fighting weapon of the immune system. 'NO is the immune system's weapon of choice against large, multicellular bodies recognized as foreign, such as transplanted organs $[51-53]$ and invasive parasites, $[54, 55]$ because the diffusion length of 'NO is particularly long. 'NO diffuses rapidly in water ($D=$ 3.3×10^{-5} cm²s⁻¹),^[56] is lipid- and watersoluble, and permeates biological membranes.[57] Its half-life in biological fluids, other than blood, is 1–20 s.^[58–60] At a $t_{1/2}$ value of $>$ 3 s, the diffusion length of 'NO is $>10^{-2}$ cm, permitting permeation through multiple cell layers.

Induction of apoptosis at high ['NO] and maintenance of high ['NO]

Nitric oxide induces apoptosis at high concentrations by S-nitrosating proteins such as GAPDH. $^{[1-3, 61]}$ iNOS expression induces immune system cytokines, including interferon- γ (IF- γ), tumor necrosis factor- α (TNF- α), and interleukin-1 β (IL- 1β).^[62] These cytokines stimulate iNOS expression in stromal cells^[63] and in tumor-infiltrating macrophages.^[64-68] Most nascent neoplasms are eliminated by apoptosis before growing and vascularizing to become large and virulent cancers.[69–73] Elimination of the nascent neoplasms fails, however, when the expression of iNOS is inhibited or when arginine, the iNOS substrate, is depleted. If a neoplasm does survive and becomes densely microvascularized, 'NO is further depleted by circulating RBCs, and apoptosis is fully arrested.

To induce apoptosis, the concentration of 'NO must be high enough to rightshift the electrochemical half-cell reaction of cysteine residues, such as those of GAPDH.^[1-3] The thermodynamic threshold concentration at which a protein is S-nitrosated is defined by the concentration of 'NO, the pH, the redox potential, and the protein concentration. Apoptosis is consequently induced only above a 'NO concentration defined by Reaction (1).

 $RSH + NO \rightarrow RS-NO + H^{+} + e^{-}$ (1)

At a sufficiently high 'NO concentration, apoptosis is extensive, and remission of tumors is observed.^[66,74,75] Maintenance of a sufficiently high 'NO concentration by the local application of a 'NO-releasing drug has resulted in the remission of tumors.^[76] Furthermore, tumorigenicity and metastasis are suppressed in iNOS-overexpressing mice.^[74,75,77-93] Also, iNOS expression and malignancy are inversely correlated in animal and human cancers, and in cancer cell lines, $[63, 94-96]$ and high 'NO concentrations produced by iNOS suppress breast tumors.^[97] Tumor development and iNOS expression are inversely correlated in human cancers;^[41, 98-105] iNOS-expressing macrophage infiltration increases the likelihood of remission in gastric,^[67,68] colorectal,^[106] and prostate cancers,^[65] and survival improves with increased tumor iNOS expression in colorectal,^[107] ovarian,^[108,109] and in non-smallcell lung cancers.[104]

Cancers associated with inadequate iNOS expression

Certain types of human tumors express little or no $iNOS.$ ^[110] This deficiency is caused by the suppression of iNOS expression at the mRNA level by TGF- $\beta1,$ ^[111] N-chlorotaurine, or N-bromotaurine. Elevated TGF- β 1 has been reported in advanced colorectal cancers.^[6] Furthermore, the level of TGF- β 1 mRNA observed in cervical smears (pap tests) correlates with the progression of cervical intra-epithelial neoplasia to cancer,^[4] and relapse of patients with breast carcinomas is known to be associated with TGF- β 1.^[7] N-chlorotaurine inhibits iNOS expression;[16, 19, 112–124] in particular, it suppresses iNOS expression in inflamed tissues[16, 19, 117–121, 123–126] and protects healthy cells from injury resulting from the overproduction of 'NO.^[19] N-chlorotaurine is formed by Reactions (2) and (3) . Reaction (2) is catalyzed by myeloperoxi $dase,$ ^[127–132] expressed in neutrophils.^[133,134] Reaction (3) proceeds without enzymatic catalysis^[21, 135, 136] at a taurine concentration of 19 mm in neutrophils.[24] Neutrophil myeloperoxidase is expressed in both human and animal brain tumors.^[17, 137] Furthermore, the (-463) G \rightarrow A point mutation in the promoter region of the myeloperoxidase gene decreases its transcription, significantly lowering the risk of lung cancer in men.[138–145]

$$
H_2O_2 + H^+ + Cl^- \rightarrow HOCl + H_2O \tag{2}
$$

$$
HOCl + NH_2CH_2CH_2SO_3^- \rightarrow
$$

ClNHCH2CH2SO3 ^þ H2O ^ð3^Þ

The analogous N-bromotaurine is produced by the eosinophil-peroxidase-catalyzed Reaction $(4)^{\left[30,146\right]}$ followed by Reaction (5), which proceeds without enzymatic catalysis in eosinophils when the taurine concentration is 15 mm. $^{[24,30]}$ Eosinophilia is characteristic of hematologic tumors, such as Hodgkin's disease,^[26] lymphomas,^[26] and certain breast carcinomas.[26, 27]

$$
H_2O_2 + H^+ + Br^- \rightarrow HOBr + H_2O \qquad \quad (4)
$$

$$
HOBr + NH2CH2CH2SO3- \rightarrow
$$

BrNHCH₂CH₂SO₃⁻ + H₂O (5)

Cancers associated with arginine depletion through overexpression of arginase

Arginase catalyzes the hydrolysis of arginine, the precursor of 'NO, to urea and ornithine. When arginase is overexpressed, tumoral arginine levels are depleted, and insufficient 'NO is produced by iNOS. Pathogens avoid being killed by iNOS-expressing macrophages through switching the macrophages from iNOS-expressing to arginase-expressing.^[147-151] Furthermore, while iNOS is active in fresh wounds, arginase is active in healing wounds, $[147, 152, 153]$ where the macrophages are again switched from iNOS-expressing to arginase-expressing.[154–157] Macrophage arginase promotes tumor cell growth and suppresses 'NO tumor cytotoxicity.^[158] Several cancer cell lines express arginase, [12,158- 162] and arginase activity is elevated in prostate,[14, 163, 164] bladder,[11] and r enal $^{[12, 13]}$ carcinomas.

Overall, inhibition of iNOS expression and arginine depletion underlie the survival of nascent neoplasms. Subsequent microvascularization makes the cancer more virulent by fully arresting apoptosis through a further decrease in the excess

'NO concentration. RBCs scavenge 'NO so rapidly that the concentration of 'NO in a volume element of tissue is defined by its distance from the nearest capillary.^[60] Circulating blood exports the residual excess 'NO from tumors through Snitrosation of GAPDH and Hb [Reaction (6)], which constitute 37–52% of the blood volume.^[59, 60, 165-176] 'NO is also stripped by oxyhemoglobin oxidation to nitrate [Reaction (7)].^[177,178] The concentration of oxyhemoglobin in the blood is \sim 2 mm, and the bimolecular rate constant for the oxidation of 'NO to nitrate is 9×10^7 M⁻¹ s⁻¹.^[34] Hence, the half-life of 'NO in blood is as short as 2×10^{-3} s,^[59] three orders of magnitude less than in other tissues.[58–60]

$$
NO + R - SH \leftrightarrow R - SNO + H^+ + e^- \qquad (6)
$$

$$
^{\bullet}NO + Hb(Fe^{II})O_2 \rightarrow Hb(Fe^{III}) + NO_3^- (7)
$$

VEGF-antibody-based anticancer drugs that slow angiogenesis were designed to starve tumors of oxygen and nutrients.^[47, 48, 179-184] Bevacizumab,^[185, 186] cetuximab,^[187] erlotinib,^[188] and trastuzumab. $[189, 190]$ which are already in use, retard the growth of cancers by preventing angiogenesis.[35–40, 191–195] However, critical analysis of the literature suggests that they act by reducing the draining of tumoral 'NO by the microvasculature.

Association of organ-specific cancers with causes of their insufficient 'NO concentrations for induction of apoptosis

Table 1 summarizes the factors that limit the 'NO concentration before vascularization. The model predicts that agents designed to sustain high local 'NO concentrations should be useful in treating cancers, particularly before microvascularization occurs. For example, locally delivered antibodies against TGF-ß1 should be of value in treating certain cervical, $[4]$ gastric,^[5] colorectal,^[6] breast,^[7,8] and lung^[9] cancers. Inhibitors of myeloperoxidase and prevention of neutrophil recruitment should be effective against some gliomas $^{[16, 17]}$ and ovarian sarcomas.[18] Inhibitors of eosinophil peroxidase and prevention of eosinophil recruitment should be useful in treating some squamous cell carcinomas of the skin,^[25] lymphomas,^[26] Hodgkin's disease.^[26] ovarian.^[26] and breast cancers.^[27] Inhibitors of arginase should be active against some bladder,^[11] renal,^[12,13] and prostate cancers.[11, 14, 15]

Table 2 lists the deductions and the facts on which they are based. Monitoring the variation of 'NO concentration in tumors should be of value in diagnosing and in determining the effectiveness of cancer treatments. Analytical methods for classifying the tumor according to iNOS or TGF- β 1 expression, the activity of arginase, myeloperoxidase, and eosinophil peroxidase, as well as the level of microvascularization are needed, as each indicates a different course of treatment.

Acknowledgements

The writing of this Essay was supported in part by the Welch Foundation (Grant F-1131).

Keywords: cancer · inhibitors · iNOS expression · nitric oxide · angiogenesis · arginine depletion

- [1] M. R. Hara, N. Agrawal, S. F. Kim, M. B. Cascio, M. Fujimuro, Y. Ozeki, M. Takahashi, J. H. Cheah, S. K. Tankou, L. D. Hester, C. D. Ferris, S. D. Hayward, S. H. Snyder, A. Sawa, Nat. Cell Biol. 2005, 7, 665–674.
- [2] M. Benhar, J. S. Stamler, Nat. Cell Biol. 2005, 7, 645–646.
- [3] M. R. Hara, S. H. Snyder, Cell. Mol. Neurobiol. 2006, 26, 525–538.
- [4] S. Baritaki, S. Sifakis, S. Huerta-Yepez, I. K. Neonakis, G. Soufla, B. Bonavida, D. A. Spandidos, Int. J. Oncol. 2007, 31, 69–79.
- [5] L. J. Hawinkels, H. W. Verspaget, W. van Duijn, J. M. van der Zon, K. Zuidwijk, F. J. Kubben, J. H. Verheijen, D. W. Hommes, C. B. Lamers, C. F. Sier, Br. J. Cancer 2007, 97, 398–404.
- [6] M. Langenskiöld, L. Holmdahl, P. Falk, E. Angenete, M. L. Ivarsson, J. Surg. Oncol. 2008, 97, 409–415.
- [7] D. L. Skerrett, E. M. Moore, D. S. Bernstein, L. Vahdat, Cancer Invest. 2005, 23, 208–214.
- [8] E. Papadopoulou, K. Anagnostopoulos, G. Tripsianis, I. Tentes, S. Kakolyris, G. Galazios, E. Sivridis, K. Simopoulos, A. Kortsaris, Neoplasma 2008, 55, 229–238.
- [9] H. G. Kang, M. H. Chae, J. M. Park, E. J. Kim, J. H. Park, S. Kam, S. I. Cha, C. H. Kim, R. W. Park, S. H. Park, Y. L. Kim, I. S. Kim, T. H. Jung, J. Y. Park, Lung Cancer 2006, 52, 1–7.
- [10] Y. Vodovotz, C. Bogdan, J. Paik, Q. W. Xie, C. Nathan, J. Exp. Med. 1993, 178, 605–613.
- [11] H. Yamanaka, T. Mayuzumi, M. Matsuoka, J. Shimazaki, K. Shida, Gann 1972, 63, 693– 700.

- Hernandez, S. Signoretti, J. Zabaleta, D. McDermott, D. Quiceno, A. Youmans, A. O'Neill, J. Mier, A. C. Ochoa, Cancer Res. 2005, 65, 3044–3048.
- [13] A. C. Ochoa, A. H. Zea, C. Hernandez, P. C. Rodriguez, Clin. Cancer Res. 2007, 13, 721s– 726s.
- G. B. Whitehurst, T. G. Pretlow, 2nd, Cancer Res. 1983, 43, 3008–3012.
- [15] A. Keskinege, S. Elgun, E. Yilmaz, Cancer Detect. Prev. 2001, 25, 76–79.
- [16] Y. Liu, M. Tonna-DeMasi, E. Park, G. Schuller-Levis, M. R. Quinn, Mol. Brain Res. 1998, 59, 189–195.
- Walker, A. Dalton, M. L. Rossi, Acta Neuropathol. 1999, 98, 349–354.
- [18] E. Oliva, J. A. Ferry, R. H. Young, J. Prat, J. R. Srigley, R. E. Scully, Am. J. Surg. Pathol. 1997, 21, 1156–1165.
- [19] J. W. Kim, C. Kim, Biochem. Pharmacol. 2005, 70, 1352–1360.

1496 <www.chemmedchem.org> © 2008 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim ChemMedChem 2008, 3, 1493 – 1499

- [20] S. T. Test, M. B. Lampert, P. J. Ossanna, J. G. Thoene, S. J. Weiss, J. Clin. Invest. 1984, 74, 1341–1349.
- [21] E. L. Thomas, M. B. Grisham, D. F. Melton, M. M. Jefferson, J. Biol. Chem. 1985, 260, 3321–3329.
- [22] S. J. Weiss, R. Klein, A. Slivka, M. Wei, J. Clin. Invest. 1982, 70, 598–607.
- [23] S. J. Weiss, M. B. Lampert, S. T. Test, Science 1983, 222, 625–628.
- [24] D. B. Learn, V. A. Fried, E. L. Thomas, J. Leukocyte Biol. 1990, 48, 174–182.
- [25] D. Lowe, C. D. Fletcher, M. P. Shaw, P. H. McKee, Histopathol. 1984, 8, 619–625.
- [26] M. Samoszuk, Histol. Histopathol. 1997, 12. 807–812.
- [27] M. K. Samoszuk, V. Nguyen, I. Gluzman, J. H. Pham, Am. J. Pathol. 1996, 148, 701–706.
- [28] R. Olszanecki, J. Marcinkiewicz, Amino Acids 2004, 27, 29–35.
- [29] A. N. Mayeno, A. J. Curran, R. L. Roberts, C. S. Foote, J. Biol. Chem. 1989, 264, 5660– 5668.
- [30] E. L. Thomas, P. M. Bozeman, M. M. Jefferson, C. C. King, J. Biol. Chem. 1995, 270, 2906–2913.
- [31] R. M. Clancy, D. Levartovsky, J. Leszczynska-Piziak, J. Yegudin, S. B. Abramson, Proc. Natl. Acad. Sci. USA 1994, 91, 3680–3684.
- [32] J. S. Stamler, D. I. Simon, J. A. Osborne, M. E. Mullins, O. Jaraki, T. Michel, D. J. Singel, J. Loscalzo, Proc. Natl. Acad. Sci. USA 1992, 89, 444–448.
- [33] M. Angelo, A. Hausladen, D. J. Singel, J. S. Stamler, Methods Enzymol. 2008, 436, 131– 168.
- [34] S. Herold, M. Exner, T. Nauser, Biochemistry 2001, 40, 3385–3395.
- [35] L. Martin, R. Schilder, J. Clin. Oncol. 2007, 25, 2894–2901.
- [36] F. J. Collinson, G. D. Hall, T. J. Perren, G. C. Jayson, Expert Rev. Anticancer Ther. 2008, 8, 21–32.
- [37] H. Bando, Breast Cancer 2007, 14, 163-173.
- [38] D. F. Hayes, K. Miller, G. Sledge, Breast 2007, 16(Suppl. 2), S17–19.
- [39] B. P. Schneider, G. W. Sledge, Jr., Nat. Clin. Pract. Oncol. 2007, 4, 181–189.
- [40] S. Sun, J. H. Schiller, Crit. Rev. Oncol. Hematol. 2007, 62, 93–104.
- [41] M. Vakkala, K. Kahlos, E. Lakari, P. Paakko, V. Kinnula, Y. Soini, Clin. Cancer Res. 2000, 6, 2408–2416.
- [42] S. L. R. Barker, H. A. Clark, S. F. Swallen, R. Kopelman, A. W. Tsang, J. A. Swanson, Anal. Chem. 1999, 71, 1767–1772.
- [43] S. R. Thom, V. Bhopale, D. Fisher, Y. Manevich, P. L. Huang, D. G. Buerk, J. Neurobiol. 2002, 51, 85–100.
- [44] A. MacKenzie, R. M. Wadsworth, Br. J. Pharmacol. 2003, 139, 1487–1497.
- [45] S. Goldstein, G. Merenyi, Methods Enzymol. 2008, 436, 49–61.
- [46] O. Gallo, E. Masini, L. Morbidelli, A. Franchi, I. Fini-Storchi, W. A. Vergari, M. Ziche, J. Natl. Cancer Inst. 1998, 90, 587–596.
- [47] Z. J. Song, P. Gong, Y. E. Wu, World J. Gastroenterol. 2002, 8, 591–595.
- [48] J. Dulak, A. Jozkowicz, Antioxid. Redox Signaling 2003, 5, 123–132.
- [49] L. Morbidelli, S. Donnini, M. Ziche, Cancer Treat. Res. 2004, 117, 155–167.
- [50] C. N. Chen, F. J. Hsieh, Y. M. Cheng, K. J. Chang, P. H. Lee, J. Surg. Oncol. 2006, 94, 226–233.
- [51] E. W. J. A. Albrecht, H. Van Goor, A. T. M. G. Tiebosch, H. Moshage, A. M. Tegzess, C. A. Stegeman, Transplantation 2000, 70, 1610-1616.
- [52] G. M. Pieper, V. Nilakantan, N. L. N. Halligan, A. K. Khanna, G. Hilton, J. Vasquez-Vivar, Biochem. J. 2005, 391. 541-547.
- [53] M. Romero, C. Garcia-Monzon, G. Clemente, M. Salcedo, E. Alvarez, P. L. Majano, R. Moreno-Otero, Liver Transplantation 2001, 7, 16–21.
- [54] X. C. Long, M. Bahgat, K. Chlichlia, A. Ruppel, Y. L. Li, J. Helminthol. 2004, 78, 47– 50.
- [55] M. Amparo Andrade, M. Siles-Lucas, J. L. Perez Arellano, C. Pou Barreto, B. Valladares, E. Espinoza, A. Muro, Nitric Oxide 2005, 13, 217–225.
- [56] T. Malinski, Z. Taha, S. Grunfeld, S. Patton, M. Kapturczak, P. Tomboulian, Biochem. Biophys. Res. Commun. 1993, 193, 1076–1082.
- [57] W. K. Subczynski, M. Lomnicka, J. S. Hyde, Free Radical Res. 1996, 24, 343–349.
- [58] G. Czapski, S. Goldstein, Free Radical Biol. Med. 1995, 19, 785–794.
- [59] X. Liu, M. J. S. Miller, M. S. Joshi, H. S. Krowicka, D. A. Clark, J. R. Lancaster, Jr., J. Biol. Chem. 1998, 273, 18 709–18 713.
- [60] D. D. Thomas, X. Liu, S. P. Kantrow, J. R. Lancaster, Jr., Proc Natl. Acad. Sci. USA 2001, 98, 355–360.
- [61] T. W. Sedlak, S. H. Snyder, JAMA J. Am. Med. Assoc. 2006, 295, 81–89.
- [62] P. A. Sherman, V. E. Laubach, B. R. Reep, E. R. Wood, Biochemistry 1993, 32, 11 600–11 605.
- [63] D. Wei, E. L. Richardson, K. Zhu, L. Wang, X. Le, Y. He, S. Huang, K. Xie, Cancer Res. 2003, 63, 3855–3859.
- [64] D. A. Arenberg, M. P. Keane, B. DiGiovine, S. L. Kunkel, S. R. Strom, M. D. Burdick, M. D. Iannettoni, R. M. Strieter, Cancer Immunol. Immunother. 2000, 49, 63–70.
- [65] S. Shimura, G. Yang, S. Ebara, T. M. Wheeler, A. Frolov, T. C. Thompson, Cancer Res. 2000, 60, 5857–5861.
- [66] M. Baratin, M. Ziol, R. Romieu, M. Kayibanda, F. Gouilleux, P. Briand, P. Leroy, H. Haddada, L. Renia, M. Viguier, J.-G. Guillet, Cancer Gene Ther. 2001, 8, 193–202.
- [67] S. Ishigami, S. Natsugoe, K. Tokuda, A. Nakajo, H. Okumura, M. Matsumoto, F. Miyazono, S. Hokita, T. Aikou, Anticancer Res. 2003, 23, 4079–4083.
- [68] S. Ohno, H. Inagawa, D. K. Dhar, T. Fujii, S. Ueda, M. Tachibana, N. Suzuki, M. Inoue, G. Soma, N. Nagasue, Anticancer Res. 2003, 23, 5015–5022.
- [69] G. P. Dunn, A. T. Bruce, H. Ikeda, L. J. Old, R. D. Schreiber, Nat. Immunol. 2002, 3, 991– 998.
- [70] M. Oren, Cell Death Differ. 2003, 10, 431– 442.
- [71] L. Römer, C. Klein, A. Dehner, H. Kessler, J. Buchner, Angew. Chem. 2006, 118, 6590– 6611; Angew. Chem. Int. Ed. 2006, 45, 6440– 6460.
- [72] G. Willimsky, T. Blankenstein, Immunol. Rev. 2007, 220, 102–112.
- [73] A. Tesniere, T. Panaretakis, O. Kepp, L. Apetoh, F. Ghiringhelli, L. Zitvogel, G. Kroemer, Cell Death Differ. 2008, 15, 3–12.
- [74] L. M. Li, R. G. Kilbourn, J. Adams, I. J. Fidler, Cancer Res. 1991, 51, 2531–2535.
- [75] K. Xie, I. J. Fidler, Cancer Metastasis Rev. 1998, 17, 55–75.
- [76] N. Katayama, K. Nakajou, H. Komori, K. Uchida, J.-i. Yokoe, N. Yasui, H. Yamamoto, T. Kai, M. Sato, T. Nakagawa, M. Takeya, T. Maruyama, M. Otagiri, J. Pharmacol. Exp. Ther. 2008, 325, 69–76.
- [77] K. Xie, S. Huang, Z. Dong, I. J. Fidler, Int. J. Oncol. 1993, 3, 1043–1048.
- [78] Z. Dong, A. H. Staroselsky, X. Qi, K. Xie, I. J. Fidler, Cancer Res. 1994, 54, 789–793.
- [79] S. Tanguay, C. D. Bucana, M. R. Wilson, I. J. Fidler, A. C. von Eschenbach, J. J. Killion, Cancer Res. 1994, 54, 5882–5888.
- [80] K. Xie, S. Huang, Z. Dong, M. Gutman, I. J. Fidler, Cancer Res. 1995, 55, 3123–3131.
- [81] K. Xie, S. Huang, Z. Dong, S. H. Juang, M. Gutman, Q. W. Xie, C. Nathan, I. J. Fidler, J. Exp. Med. 1995, 181, 1333–1343.
- [82] K. Xie, Z. Dong, I. J. Fidler, J. Leukocyte Biol. 1996, 59, 797–803.
- [83] K. Xie, S. Huang, Y. Wang, P. J. Beltran, S. H. Juang, Z. Dong, J. C. Reed, T. J. McDonnell, D. J. McConkey, I. J. Fidler, Cancer Immunol. Immunother. 1996, 43, 109–115.
- [84] S. H. Juang, K. Xie, L. Xu, Y. Wang, J. Yoneda, I. J. Fidler, Cancer Biother. Radiopharm. 1997, 12, 167–175.
- [85] K. Xie, D. Bielenberg, S. Huang, L. Xu, T. Salas, S. H. Juang, Z. Dong, I. J. Fidler, Clin. Cancer Res. 1997, 3, 2283–2294.
- [86] K. Xie, S. Huang, Z. Dong, S. H. Juang, Y. Wang, I. J. Fidler, J. Natl. Cancer Inst. 1997, 89, 421–427.
- [87] K. Xie, Y. Wang, S. Huang, L. Xu, D. Bielenberg, T. Salas, D. J. McConkey, W. Jiang, I. J. Fidler, Oncogene 1997, 15, 771–779.
- [88] I. Eue, R. Kumar, Z. Dong, J. J. Killion, I. J. Fidler, J. Immunother. 1998, 21, 340–351.
- [89] S. H. Juang, K. Xie, L. Xu, Q. Shi, Y. Wang, J. Yoneda, I. J. Fidler, Hum. Gene Ther. 1998, 9, 845–854.
- [90] L. Xu, K. Xie, I. J. Fidler, Hum. Gene Ther. 1998, 9, 2699–2708.
- [91] C. J. Bruns, H. Shinohara, M. T. Harbison, D. W. Davis, G. Nelkin, J. J. Killion, D. J. McConkey, Z. Dong, I. J. Fidler, Cancer Res. 2000, 60, 2–7.
- [92] H. Shinohara, C. D. Bucana, J. J. Killion, I. J. Fidler, J. Immunother. 2000, 23, 321–331.
- [93] S. Ozawa, W. Lu, C. D. Bucana, H. O. Kanayama, H. Shinohara, I. J. Fidler, Z. Dong, Int. J. Oncol. 2003, 22, 977–984.
- [94] Q. Shi, Q. Xiong, B. Wang, X. Le, N. A. Khan, K. Xie, Cancer Res. 2000, 60, 2579–2583.
- [95] X. Le, D. Wei, S. Huang, J. R. Lancaster, Jr., K. Xie, Proc. Natl. Acad. Sci. USA 2005, 102, 8758–8763.
- [96] J. R. Lancaster, Jr., K. Xie, Cancer Res. 2006, 66, 6459–6462.
- [97] A. Pance, Future Oncol. 2006, 2, 275–288.
- [98] S. Reveneau, L. Arnould, G. Jolimoy, S. Hilpert, P. Lejeune, V. Saint-Giorgio, C. Beli-

chard, J.-F. Jeannin, Lab. Invest. 1999, 79, 1215–1225.

- [99] W. Tschugguel, T. Pustelnik, H. Lass, M. Mildner, W. Weninger, C. Schneeberger, B. Jansen, E. Tschachler, T. Waldhor, J. C. Huber, H. Pehamberger, Br. J. Cancer 1999, 79, 1609–1612.
- [100] W. Tschugguel, C. Schneeberger, G. Unfried, K. Czerwenka, W. Weninger, M. Mildner, D. M. Gruber, M. O. Sator, T. Waldhor, J. C. Huber, Breast Cancer Res. Treat. 1999, 56, 143–151.
- [101] G. Kong, E. K. Kim, W. S. Kim, K. T. Lee, Y. W. Lee, J. K. Lee, S. W. Paik, J. C. Rhee, J. Gastroenterol. Hepatol. 2002, 17, 914–921.
- [102] G. Kong, E. K. Kim, W. S. Kim, Y. W. Lee, J. K. Lee, S. W. Paik, J. C. Rhee, K. W. Choi, K. T. Lee, Int. J. Pancreatol. 2002, 29, 133–140.
- [103] A. Jayasurya, S. T. Dheen, W. M. Yap, N. G. Tan, Y. K. Ng, B. H. Bay, Int. J. Radiat. Oncol. Biol. Phys. 2003, 56, 837–845.
- [104] A. Puhakka, V. Kinnula, U. Napankangas, M. Saily, P. Koistinen, P. Paakko, Y. Soini, Apmis 2003, 111, 1137–1146.
- [105] J. Mazibrada, M. Ritta, M. Mondini, M. De Andrea, B. Azzimonti, C. Borgogna, M. Ciotti, A. Orlando, N. Surico, L. Chiusa, S. Landolfo, M. Gariglio, Gynecol. Oncol. 2008, 108, 112–120.
- [106] Y. Funada, T. Noguchi, R. Kikuchi, S. Takeno, Y. Uchida, H. E. Gabbert, Oncol. Rep. 2003, 10, 309–313.
- [107] K. M. Ropponen, J. K. Kellokoski, P. K. Lipponen, M. J. Eskelinen, L. Alanne, E. M. Alhava, V. M. Kosma, Scand. J. Gastroenterol. 2000, 35, 1204–1211.
- [108] M. R. Raspollini, G. Amunni, A. Villanucci, V. Boddi, G. Baroni, A. Taddei, G. L. Taddei, Gynecol. Oncol. 2004, 92, 806–812.
- [109] M. A. Anttila, K. Voutilainen, S. Merivalo, S. Saarikoski, V. M. Kosma, Gynecol. Oncol. 2007, 105, 97–103.
- [110] J. A. Crowell, V. E. Steele, C. C. Sigman, J. R. Fay, Mol. Cancer Ther. 2003, 2, 815–823.
- [111] T. Mitani, M. Terashima, H. Yoshimura, Y. Nariai, Y. Tanigawa, Nitric Oxide 2005, 13, 78–87.
- [112] Y. Y. Lin, C. E. Wright, M. Zagorski, K. Nakanishi, Biochim. Biophys. Acta Mol. Cell Res. 1988, 969, 242–248.
- [113] L. A. Marquez, H. B. Dunford, J. Biol. Chem. 1994, 269, 7950–7956.
- [114] G. Schuller-Levis, M. R. Quinn, C. Wright, E. Park, Adv Exp Med. Biol. 1994, 359, 31–39.
- [115] D. R. Ramos, M. Victoria Garcia, L. M. Canle, J. Arturo Santaballa, P. G. Furtmuller, C. Obinger, Arch. Biochem. Biophys. 2007, 466, 221–233.
- [116] E. Park, M. R. Quinn, C. E. Wright, G. Schuller-Levis, J. Leukocyte Biol. 1993, 54, 119– 124.
- [117] G. B. Schuller-Levis, W. R. Levis, M. Ammazzalorso, A. Nosrati, E. Park, Infect. Immun. 1994, 62, 4671–4674.
- [118] E. Park, G. Schuller-Levis, M. R. Quinn, J. Immunol. 1995, 154, 4778–4784.
- [119] C. Kim, E. Park, M. R. Quinn, G. Schuller-Levis, Immunopharmacology 1996, 34, 89– 95.

- [120] J. Marcinkiewicz, A. Grabowska, J. Bereta, T. Stelmaszynska, J. Leukocyte Biol. 1995, 58, 667–674. [121] E. Park, G. Schuller-Levis, J.-H. Jia, M. R.
- Quinn, J. Leukocyte Biol. 1997, 61, 161–166. [122] M. Barua, Y. Liu, M. R. Quinn, J. Immunol.
- 2001, 167, 2275–2281. [123] V. Serban, Y. Liu, M. R. Quinn, Adv. Exp. Med.
- Biol. 2003, 526, 357–364.
- [124] M. R. Quinn, M. Barua, Y. Liu, V. Serban, Adv. Exp. Med. Biol. 2003, 526, 341–348.
- [125] E. Park, J. Jia, M. R. Ouinn, G. Schuller-Levis, Clin. Immunol. 2002, 102, 179–184.
- [126] C. Kim, S. Choi Hyung, W. Kim Jun, Adv. Exp. Med. Biol. 2006, 583, 493–498.
- [127] E. L. Thomas, M. Fishman, J. Biol. Chem. 1986, 261, 9694–9702.
- [128] A. J. Kettle, C. C. Winterbourn, Methods Enzymol. 1994, 233, 502–512.
- [129] E. L. Thomas, Infect. Immun. 1979, 23, 522-531.
- [130] J. M. Albrich, C. A. McCarthy, J. K. Hurst, Proc. Natl. Acad. Sci. USA 1981, 78, 210–214. [131] C. C. Winterbourn, A. J. Kettle, Free Radical
- Biol. Med. 2000, 29, 403–409. [132] S. J. Klebanoff, J. Leukocyte Biol. 2005, 77.
- 598–625.
- [133] E. L. Thomas, R. I. Lehrer, R. F. Rest, Rev. Infect. Dis. 1988, 10 (Suppl. 2), S450–456.
- [134] C.C. Winterbourn, Toxicology 2002, 181-182, 223–227.
- [135] C. Cunningham, K. F. Tipton, H. B. F. Dixon, Biochem. J. 1998, 330, 939–945.
- [136] A. V. Peskin, R. G. Midwinter, D. T. Harwood, C. C. Winterbourn, Free Radical Biol. Med. 2004, 37, 1622–1630.
- [137] M. Cao, C. Eshoa, C. Schultz, J. Black, Y. Zu, C. C. Chang, Arch. Pathol. Lab. Med. 2007, 131, 301–305.
- [138] I. Cascorbi, S. Henning, J. Brockmoller, J. Gephart, C. Meisel, J. M. Muller, R. Loddenkemper, I. Roots, Cancer Res. 2000, 60, 644–649.
- [139] A. Feyler, A. Voho, C. Bouchardy, K. Kuokkanen, P. Dayer, A. Hirvonen, S. Benhamou, Cancer Epidemiol. Biomarkers Prev. 2002, 11, 1550–1554.
- [140] O. H. Kantarci, T. G. Lesnick, P. Yang, R. L. Meyer, D. D. Hebrink, C. T. McMurray, B. G. Weinshenker, Mayo Clin. Proc. 2002, 77, 17– 22.
- [141] W. Lu, D. Xing, J. Qi, W. Tan, X. Miao, D. Lin, Int. J. Cancer 2002, 102, 275–279.
- [142] M. B. Schabath, M. R. Spitz, W. K. Hong, G. L. Delclos, W. F. Reynolds, G. B. Gunn, L. W. Whitehead, X. Wu, Lung Cancer 2002, 37, 35–40.
- [143] X. Wu, M. B. Schabath, M. R. Spitz, Methods Mol. Med. 2003, 75, 121–133.
- [144] C. Kiyohara, K. Yoshimasu, K. Takayama, Y. Nakanishi, Genet. Med. 2005, 7, 463–478.
- [145] J. H. Park, J. M. Park, E. J. Kim, S. I. Cha, E. B. Lee, C. H. Kim, S. Kam, T. H. Jung, J. Y. Park, Cancer Detect. Prev. 2006, 30, 257–261.
- [146] S. J. Weiss, S. T. Test, C. M. Eckmann, D. Roos, S. Regiani, Science 1986, 234, 200– 203.
- [147] M. J. Rauh, L. M. Sly, J. Kalesnikoff, M. R. Hughes, L. P. Cao, V. Lam, G. Krystal, Biochem. Soc. Trans. 2004, 32, 785–788.
- [148] M. Hesse, M. Modolell, A. C. La Flamme, M. Schito, J. M. Fuentes, A. W. Cheever, E. J.
- Pearce, T. A. Wynn, J. Immunol. 2001, 167, 6533–6544.
- [149] A. P. Gobert, Y. Cheng, J. Y. Wang, J. L. Boucher, R. K. Iyer, S. D. Cederbaum, R. A. Casero, Jr., J. C. Newton, K. T. Wilson, J. Immunol. 2002, 168, 4692–4700.
- [150] A. L. Pauleau, R. Rutschman, R. Lang, A. Pernis, S. S. Watowich, P. J. Murray, J. Immunol. 2004, 172, 7565–7573.
- [151] K. W. Raines, T. J. Kang, S. Hibbs, G. L. Cao, J. Weaver, P. Tsai, L. Baillie, A. S. Cross, G. M. Rosen, Infect. Immun. 2006, 74, 2268–2276.
- [152] J. E. Albina, C. D. Mills, W. L. Henry, Jr., M. D. Caldwell, J. Immunol. 1990, 144, 3877–3880.
- [153] J. D. Shearer, J. R. Richards, C. D. Mills, M. D. Caldwell, Am. J. Physiol. 1997, 272, E181– 190.
- [154] B. Benninghoff, V. Lehmann, H. P. Eck, W. Droge, Int. Immunol. 1991, 3, 413–417.
- [155] M. R. Schaffer, U. Tantry, R. A. van Wesep, A. Barbul, J. Surg. Res. 1997, 71, 25–31.
- [156] S. Gordon, Nat. Rev. Immunol. 2003, 3, 23– 35.
- [157] J. E. Albina, E. J. Mahoney, J. M. Daley, D. E. Wesche, S. M. Morris, Jr., J. S. Reichner, Shock 2005, 23, 168–172.
- [158] C.-I. Chang, J. C. Liao, L. Kuo, Cancer Res. 2001, 61, 1100–1106.
- [159] Y. Honma, Y. Fujita, J. Okabe-Kado, T. Kasukabe, M. Hozumi, Cancer Lett. 1980, 10, 287–292.
- [160] R. Singh, S. Pervin, A. Karimi, S. Cederbaum, G. Chaudhuri, Cancer Res. 2000, 60, 3305– 3312.
- [161] O. Margalit, L. Eisenbach, N. Amariglio, N. Kaminski, A. Harmelin, R. Pfeffer, M. Shohat, G. Rechavi, R. Berger, Br. J. Cancer 2003, 89, 314–319.
- [162] B. Melichar, W. Hu, R. Patenia, K. Melicharova, S. T. Gallardo, R. Freedman, J. Transl. Med. 2003, 1, 5.
- [163] T. G. Pretlow II, B. E. Harris, E. L. Bradley, Jr., A. J. Bueschen, K. L. Lloyd, T. P. Pretlow, Cancer Res. 1985, 45, 442–446.
- [164] A. Z. Vafa, P. K. Grover, T. G. Pretlow, M. I. Resnick, Urology 1993, 42, 138–143.
- [165] T. S. Hakim, K. Sugimori, E. M. Camporesi, G. Anderson, Physiol. Meas 1996, 17, 267–277.
- [166] A. R. Butler, I. L. Megson, P. G. Wright, Biochim. Biophys. Acta Gen. Subj. 1998, 1425, 168–176.
- [167] R. Rossi, D. Barra, A. Bellelli, G. Boumis, S. Canofeni, P. Di Simplicio, L. Lusini, S. Pascarella, G. Amiconi, J. Biol. Chem. 1998, 273, 19 198–19 206.
- [168] M. W. Vaughn, L. Kuo, J. C. Liao, Am. J. Physiol. 1998, 274, H1705–H1714.
- [169] K.-T. Huang, T. H. Han, D. R. Hyduke, M. W. Vaughn, H. Van Herle, T. W. Hein, C. Zhang, L. Kuo, J. C. Liao, Proc. Natl. Acad. Sci. USA 2001, 98, 11 771–11 776.
- [170] M. S. Joshi, T. B. Ferguson, Jr., T. H. Han, D. R. Hyduke, J. C. Liao, T. Rassaf, N. Bryan, M. Feelisch, J. R. Lancaster, Jr., Proc. Natl. Acad. Sci. USA 2002, 99, 10 341–10 346.
- [171] X. Liu, A. Samouilov, J. R. Lancaster, Jr., J. L. Zweier, J. Biol. Chem. 2002, 277, 26 194– 26 199.
- [172] T. H. Han, D. R. Hyduke, M. W. Vaughn, J. M. Fukuto, J. C. Liao, Proc. Natl. Acad. Sci. USA 2002, 99, 7763–7768.

- [173] T. H. Han, E. Qamirani, A. G. Nelson, D. R. Hyduke, G. Chaudhuri, L. Kuo, J. C. Liao, Proc. Natl. Acad. Sci. USA 2003, 100, 12 504– 12 509.
- [174] I. Azarov, K. T. Huang, S. Basu, M. T. Gladwin, N. Hogg, D. B. Kim-Shapiro, J. Biol. Chem. 2005, 280, 39 024–39 032.
- [175] J. Sandmann, K. S. Schwedhelm, D. Tsikas, FEBS Lett. 2005, 579, 4119–4124.
- [176] K. T. Huang, Z. Huang, D. B. Kim-Shapiro, Nitric Oxide 2007, 16, 209–216.
- [177] L. J. Ignarro, J. M. Fukuto, J. M. Griscavage, N. E. Rogers, R. E. Byrns, Proc. Natl. Acad. Sci. USA 1993, 90, 8103–8107.
- [178] P. R. Gardner, A. M. Gardner, W. T. Brashear, T. Suzuki, A. N. Hvitved, K. D. R. Setchell, J. S. Olson, J. Inorg. Biochem. 2006, 100, 542– 550.
- [179] R. D. Leek, N. C. Hunt, R. J. Landers, C. E. Lewis, J. A. Royds, A. L. Harris, J. Pathol. 2000, 190, 430–436.
- [180] M. C. Duyndam, M. C. Hilhorst, H. M. Schluper, H. M. Verheul, P. J. van Diest, G. Kraal, H. M. Pinedo, E. Boven, Am. J. Pathol. 2002, 160, 537–548.
- [181] F. Cianchi, C. Cortesini, O. Fantappie, L. Messerini, N. Schiavone, A. Vannacci, S. Nistri, I.

Sardi, G. Baroni, C. Marzocca, F. Perna, R. Mazzanti, P. Bechi, E. Masini, Am. J. Pathol. 2003, 162, 793–801.

- [182] R. Bartoletti, T. Cai, G. Nesi, I. Sardi, M. Rizzo, Oncol. Rep. 2005, 14, 251–255.
- [183] S. Tsutsui, K. Yasuda, K. Suzuki, K. Tahara, H. Higashi, S. Era, Oncol. Rep. 2005, 14, 425– 431.
- [184] E. R. Camp, A. Yang, W. Liu, F. Fan, R. Somcio, D. J. Hicklin, L. M. Ellis, Clin. Cancer Res. 2006, 12, 2628–2633.
- [185] J. R. Jett, S. E. Schild, R. L. Keith, K. A. Kesler, Chest 2007, 132(Suppl. 3), 266S-276S.
- [186] M. A. Socinski, R. Crowell, T. E. Hensing, C. J. Langer, R. Lilenbaum, A. B. Sandler, D. Morris, Chest 2007, 132(Suppl. 3), 277S– 289S.
- [187] J. Bernier, J. Bonner, J. B. Vermorken, R. J. Bensadoun, R. Dummer, J. Giralt, G. Kornek, A. Hartley, R. Mesia, C. Robert, S. Segaert, K. K. Ang, Ann. Oncol. 2007, 19, 142–149.
- [188] J. Noble, P. M. Ellis, J. A. Mackay, W. K. Evans, J. Thorac. Oncol. 2006, 1, 1042–1058.
- [189] R. W. Carlson, E. Brown, H. J. Burstein, W. J. Gradishar, C. A. Hudis, C. Loprinzi, E. P. Mamounas, E. A. Perez, K. Pritchard, P. Ravdin, A. Recht, G. Somlo, R. L. Theriault, E. P.

Winer, A. C. Wolff, J. Natl. Compr. Cancer Network 2006, 4 (Suppl. 1), S1–26.

- [190] R. W. Carlson, S. J. Moench, M. E. Hammond, E. A. Perez, H. J. Burstein, D. C. Allred, C. L. Vogel, L. J. Goldstein, G. Somlo, W. J. Gradishar, C. A. Hudis, M. Jahanzeb, A. Stark, A. C. Wolff, M. F. Press, E. P. Winer, S. Paik, B. M. Ljung, J. Natl. Compr. Cancer Network 2006, 4 (Suppl. 3), S1–22; quiz S23–24.
- [191] D. Liao, R. S. Johnson, Cancer Metastasis Rev. 2007, 26, 281–290.
- [192] R. K. Jain, E. di Tomaso, D. G. Duda, J. S. Loeffler, A. G. Sorensen, T. T. Batchelor, Nat. Rev. Neurosci. 2007, 8, 610–622.
- [193] S. B. Fox, D. G. Generali, A. L. Harris, Breast Cancer Res. 2007, 9, 216.
- [194] S. Banerjee, M. Dowsett, A. Ashworth, L. A. Martin, Nat. Clin. Pract. Oncol. 2007, 4, 536– 550.
- [195] A. L. Harzstark, C. J. Ryan, Expert Rev. Anticancer Ther. 2008, 8, 259–268.

Received: July 30, 2008 Published online on August 29, 2008